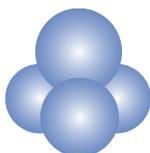


# Product Specification Sheet

<b>Product Name</b>	<b>ABCA7 Human Induced Pluripotent Stem Cells (Episomal, HFF)</b>
<b>Description</b>	Footprint-free human iPS (induced pluripotent stem) cell line with ATP-binding cassette sub-family A member 7 (ABCA7) deficiency was derived using genome editing methods to introduce a deletion and frame shift in the ABCA7 gene. Sequencing results confirmed that this isogenic iPSC line has a stable homozygous 82 bp deletion in the exon 8 of ABCA7 gene. When cultured under standard human ES cell culture conditions, the morphology of human iPS cells is identical to that of human ES cells. The cells also express the pluripotency markers TRA-1-60, SSEA-3 and Oct4, and demonstrate strong endogenous alkaline phosphatase activity.
<b>Catalog Number</b>	iPS21
<b>Size</b>	> 5x10 <sup>5</sup> cells/vial
<b>Shipping</b>	Dry ice
<b>Storage and Stability</b>	Store in vapor phase of liquid nitrogen immediately upon receipt. This product is stable for 6 months when stored as directed.
<b>Quality Control</b>	Human iPS cells were grown in feeder free conditions with mTeSR1 medium. Each lot of human iPS cells is tested for growth and viability following recovery from cryopreservation. In addition, each lot is tested for expression of TRA-1-60 and OCT4, as well as the activity of alkaline phosphatase.
<b>Safety Precaution</b>	<b>Alstem highly recommends that protective gloves, a lab coat, and a full-face mask are always worn when handling frozen vials.</b> It is important to note that some liquid nitrogen can leak into the vials when submersed in liquid nitrogen. Upon thawing, the liquid nitrogen returns to the gas phase, resulting in excessive pressure within the vial that can cause the vial to explode or expel the cap with dangerous force.
<b>Restricted Use</b>	For Research Use Only. Not for use in diagnostic or therapeutic procedures.



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# Overview

ABCA7 protein is a member of the superfamily of ATP-binding cassette (ABC) transporters which regulate the lipid metabolism in the central nervous system, as well as peripheral tissues. Genome-wide association studies (GWAS) identified several ABCA7 variants as novel risk factors for Alzheimer’s disease (AD). In vitro studies indicate over-expression of ABCA7 results in a significant decrease in amyloid- $\beta$  ( $A\beta$ ) production, while a more rapid uptake of amyloid precursor protein (APP) in ABCA7 knockout cells is mechanistically consistent with an increased  $A\beta$  production. It was therefore suggested that ABCA7 protein directly impacts amyloid- $\beta$  pathology by altering APP trafficking and substrate availability.

Alstem’s footprint-free human iPS (induced pluripotent stem) cell line (cat# iPS21) carries ABCA7 KO mutation in both alleles. Sequencing results confirmed that this isogenic iPSC line has a stable homozygous 82 bp deletion in the exon 8 of ABCA7 gene. The morphology of this human iPSC cell line is identical to that of human ES cells. The cells also express the pluripotency markers TRA-1-60, SSEA-3 and OCT4, and demonstrate strong endogenous alkaline phosphatase activity.

Alstem’s human iPS cells are a good model for studying AD-related pathologies in neurons induced specifically by ABCA7, as well as for drug discovery.

## Data Analysis



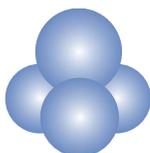
**Figure 1.** The sequencing results showed this iPSC line (cat# iPS21) had an 82-bp deletion in the exon 8 of ABCA7 gene in both alleles.

# Protocol

## Human iPS Cell Culture

### OVERVIEW

This protocol can be used for culturing human iPS cells. Footprint-free human iPS cells were generated by transient introduction of episomal plasmids encoding the human transcription factors into human peripheral blood mononuclear cells. The cells were derived using morphological selection criteria without the use of fluorescent markers or drug selection. When cultured under standard human ES cell culture conditions, the



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morphology of footprint-free human iPS cells is identical to that of human ES cells. The cells also express the pluripotency markers TRA-1-60 and Oct4, and demonstrate a strong endogenous AP activity.

## PROCEDURE

### I. Feeder free culture conditions

#### Preparation of feeder-free medium

1. Thaw mTeSR1 5X Supplement (Cat.no. 05850, STEMCELL Technologies) at room temperature or overnight at 4°C.
2. Add the 100 mL of thawed 5X Supplement to 400 mL Basal Medium for a total volume of 500 mL aseptically. Mix well and filter through a 0.2 µm, low-protein binding filter, if desired.
3. Aliquot into appropriate amount according to usage and store the aliquots at 4°C.

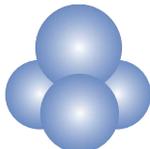
#### Coating plates with Matrigel

Matrigel (Cat.no. 354277, BD) should be aliquoted and stored at -80°C for long-term use.

1. Thaw matrigel on ice until liquid. Dilute matrigel 1:30 to 1:50 with pre-chilled KO DMEM/F12.
2. Immediately use the diluted matrigel solution to coat tissue culture-treated plates. For a 6-well plate, use 1 mL of diluted matrigel solution per well, and swirl the plate to spread the matrigel solution evenly across the surface.
3. Let the coated plate stand for 2 h at 37°C or overnight at 4°C. If plate has been stored at 4°C, allow the plate to incubate at 37°C for at least 30 minutes before removing the matrigel solution.

#### Thawing cryopreserved human iPS cells

1. Quickly thaw the human iPS cells in a 37°C water bath by gently shaking the cryovial continuously until half thawed. Remove the cryovial from the water bath and spray with 70% ethanol to sterilize.
2. Transfer the contents of the cryovial to a 15 mL conical tube. Add 5 mL warm mTeSR1 dropwise to the tube, gently mixing as the medium is added.
3. Centrifuge cells at 200 x g for 5 minutes at room temperature.
4. After centrifugation, aspirate the medium from 15 mL tube. Gently resuspend the cell pellet in 2 mL pre-warmed mTeSR1 with 5 µM ROCK inhibitor media. Avoid overpipetting to ensure maintaining the cells as small cell clumps.
5. Remove the matrigel solution from a coated tissue culture 6-well plate. Transfer the cell suspension to the matrigel coated 6-well plate.
6. Place the plate into the 37°C incubator and move the plate in quick side to side, forward to back motions to evenly distribute the cells within the wells. Culture the cells at 37°C, with 5% CO<sub>2</sub> and 95% humidity.
7. Change medium daily. Check for undifferentiated colonies that are ready to passage when colonies are big enough (approximately 7-10 days after thawing).



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### **Passaging human iPS cells grown under feeder-free conditions**

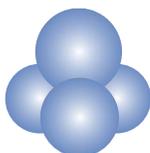
1. Use a microscope to identify regions of differentiation. Mark the differentiated colonies using lens marker on the bottom of the plate.
  2. Remove regions of differentiation by scraping with a pipette tip or by aspiration.
  3. Aspirate medium from the human iPS cell culture and rinse with DPBS (2 mL/well).
  4. Add 1 mL per well of EZStem Enzyme-Free Stem Cell Dissociation Solution (cat. no. M100, Alstem) and incubate at 37°C for 2-3 minutes.
- NOTE: Addition of 0.5 mL per well of Accutase (Cat.no. SCR005, Millipore, diluted 1:1 with DPBS before use) and incubate at 37°C for 1 minute can also be used for dissociation of cells.
5. Remove EZStem Enzyme-Free Stem Cell Dissociation Solution or Accutase by pipetting the solution out of the well, gently rinse each well 2 - 3 times with 2 mL of DMEM/F-12 per well and transfer the detached cell aggregates to a 15 mL conical tube.
  6. Add 2 mL/well mTeSR1 and scrape colonies off with a cell scraper. Transfer the detached cell aggregates to a 15 mL conical tube.
  7. Rinse the well with an additional 2 mL mTeSR1 to collect any remaining cell aggregates. Add the remaining cell suspension to the 15 mL tube.
  8. Centrifuge the 15 mL tube containing the cell suspension at 200 x *g* for 5 minutes at room temperature.
  9. Aspirate the supernatant. Resuspend pellet in mTeSR1 containing 5 µM ROCK inhibitor by gently pipetting to ensure that the cells are maintained as aggregates.
  10. Plate the human iPS cell aggregates with mTeSR1 in a new plate coated with matrigel. (Remove matrigel solution before plating). *If the colonies are at an optimal density, the cells can be split every 5 - 7 days using 1:3 to 1:6 ratios.*
  11. Place the plate into the 37°C incubator and move the plate in quick side to side and forward to back motions to evenly distribute the clumps within the wells. Culture the cells at 37°C, with 5% CO<sub>2</sub> and 95% humidity.
  12. Change the cell culture medium daily.

### **Cryopreserving human iPS cells**

1. Prepare EZStem freezing medium (Cat.no. M050, ALSTEM) on ice.
2. Perform steps 1-8 from **Passaging human iPS cells grown under feeder-free conditions**
3. Gently aspirate the supernatant and loosen the cell pellet by tapping the bottom of the tube.
4. Gently resuspend the pellet in freezing medium, taking care to leave the clumps larger than those that would normally be done for passaging.
5. Transfer 1 mL of cells in freezing medium into each labeled cryogenic vial.
6. Place vials into a freezing container and place the container at -80°C overnight.
7. Transfer to a liquid nitrogen tank next day.

### **IMPORTANT NOTICE**

Store the vials at vapor phase of liquid nitrogen immediately upon receipt.



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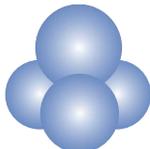
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## **WARNING**

**Do not use cryogenic vials for storage in the liquid phase of liquid nitrogen. Such use may cause entrapment of liquid nitrogen inside the vial and lead to pressure buildup resulting in possible explosion or biohazard release. Use appropriate safety procedures which are outlined by the ATCC when handling and disposing of vials. ALSTEM highly recommends that protective gloves and clothing always be used and a full face mask always be worn when handling frozen vials.**



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